Hindawi Evidence-Based Complementary and Alternative Medicine Volume 2020, Article ID 2580259, 12 pages https://doi.org/10.1155/2020/2580259



Review Article

Ethnobotanical Description and Biological Activities of Senna alata

Oluwole Solomon Oladeji (p), 1,2 Funmilayo Enitan Adelowo, 2 Abimbola Peter Oluyori, 1 and Deborah Temitope Bankole (p) 1

¹Natural Products Research Unit, Industrial Chemistry Programme, Department of Physical Sciences, Faculty of Pure and Applied Sciences, Landmark University, Omu-Aran, Nigeria

Correspondence should be addressed to Oluwole Solomon Oladeji; oladeji.oluwole@lmu.edu.ng

Received 24 September 2019; Revised 16 January 2020; Accepted 27 January 2020; Published 21 February 2020

Academic Editor: Marisa Colone

Copyright © 2020 Oluwole Solomon Oladeji et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Senna alata is a medicinal herb of Leguminosae family. It is distributed in the tropical and humid regions. The plant is traditionally used in the treatment of typhoid, diabetes, malaria, asthma, ringworms, tinea infections, scabies, blotch, herpes, and eczema. The review is aimed at unveiling the ethnobotanical description and pharmacological activities of S. alata. Different parts of the plant are reported in folk medicine as therapeutic substances for remediation of diverse diseases and infections. The extracts and isolated compounds displayed pronounced pharmacological activities. Display of antibacterial, antioxidant, antifungal, dermatophytic, anticancer, hepatoprotective, antilipogenic, anticonvulsant, antidiabetic, antihyperlipidemic, antimalarial, anthelmintic, and antiviral activities could be due to the array of secondary metabolites such as tannins, alkaloids, flavonoids, terpenes, anthraquinone, saponins, phenolics, cannabinoid alkaloids, 1,8-cineole, caryophyllene, limonene, α -selinene, β -caryophyllene, germacrene D, cinnamic acid, pyrazol-5-ol, methaqualone, isoquinoline, quinones, reducing sugars, steroids, and volatile oils present in different parts of the plant. The review divulges the ethnobotanical and pharmacological activities of the plant and also justifies the ethnomedical claims. The significant medicinal value of this plant necessitates a scientific adventure into the bioactive metabolites which constitute various extracts.

1. Introduction

The primary and safest therapeutic approach since prehistoric time is herbal medicine which has displayed significant role in primary health care development [1]. The World Health organization (WHO) appraised that 80% of inhabitants in developing countries basically rely on herbal medicines [2, 3]. Recently, it was discovered that one-third of the commonly used drugs are obtained from natural sources. This has led to the documentation of about 40,000–70,000 medicinal plant species with outstanding therapeutic potentials [4].

The scientific appraisal of the pharmacological activities of herbal plants revealed about 200,000 phytochemicals.

These compounds contribute to the apparent medicinal activities displayed by plants and invariably justify the involvement of natural products in the development of novel drugs [5]. Despite these achievements, the use of herbal drugs was not universally accepted in contemporary medicine due to lack of scientific evidence and proper documentation. However, the significance of herbs in pharmacology has necessitated the provision of scientific facts on bioactive compounds and pharmacological assays of plants [6, 7].

Medicinal plants with immense biological applications have been viewed as the principal sources of therapeutic substances which could lead to novel therapeutic compounds. The search for these compounds from medicinal

²Department of Pure and Applied Chemistry, Faculty of Pure and Applied Sciences, Ladoke Akintola University of Technology, PMB 4000, Ogbomoso, Nigeria

plants usually ends in the isolation of novel compounds and, eventually, drug developments [8]. Several medicinal plants with diverse interesting pharmacophore have been scientifically investigated, and one of these plants is *Senna alata*.

S. alata, also known as Cassia alata, is a widely distributed herb of the Leguminosae family. It is commonly known as candle bush, craw-craw plant, acapulo, ringworm bush, or ringworm plant. The plant is commonly found in Asia and Africa, and has many local names [9]. It has arrays of bioactive chemical compounds. Some of the reported chemical constituents are phenolics (rhein, chrysaphanol, kaempferol, aloeemodin, and glycosides), anthraquinones (alatinone and alatonal), fatty acids (oleic, palmitic, and linoleic acids), steroids, and terpenoids (sitosterol, stigmasterol, and campesterol) [10, 11]. These secondary metabolites are reported to display numerous biological activities [12–17].

The flower, root, leaves, seed, and bark displayed diverse biological activities [18–20]. These pharmacological activities include antimicrobial [21–23], antifungal [24], anticryptococcus [25], antibacterial [26, 27], antitumor [28], anti-inflammatory [29, 30], antidiabetic [31], antioxidant [18], wound healing [32], and antihelmintic activities [33]. In recent times, the outbreak of drug-resistant diseases has led to several health issues. In an attempt to resolve these issues, pharmacological research has been tailored towards the discovery of innovative, potent, and safe drugs from natural compounds. This review appraises the ethnobotanical and pharmacological activities of S. *alata*, thus justifying diverse traditional applications of the plant.

2. Review Methodology

Relevant documents were obtained from major scientific catalogues such as Pubmed, Google Scholar, EBSCO, Sci-Finder, Scopus, Medline, and Science Direct. Many publications sites were queried to procure information on ethnobotanical description and pharmacological activities of *S. alata*.

3. Ethnobotanical Description

3.1. Description and Classification. S. alata (L.) Roxb. is a flowering shrub of the Fabaceae family. It has the name "candle bush" owing to the framework of its inflorescences [34]. It is an annual and occasionally biannual herb, with an average height of 1 to 4 m, burgeoning in sunlit and humid zones. The leaves are oblong, with 5 to 14 leaflet sets, robust petioles (2 to 3 mm), caduceus bracts (2×3 by 1×2 cm), and dense flowers (2×50 by 3×4 cm). Zygomorphic flowers have bright yellow color with 7 stamens and a pubertal ovary. The fruit exists as a 10 to 16×1.5 cm tetragonal pod, thick, flattened wings, brown when ripe with many diamond-shaped brown seeds. It is propagated by seeds and dispersed to about 1,500 m above the sea level [35, 36].

Taxonomically, *S. alata* is classified as kingdom: Plantae; order: Fabales; family: Fabaceae; subfamily: Caesalpinioideae; tribe: Cassieae; subtribe: Cassiinae; genus: Senna; species: *S. alata*.

3.2. Geographical Distribution. S. alata is widely distributed in Ghana, Brazil, Australia, Egypt, India, Somalia, Sri Lanka, and all over Africa [37]. It is an ornamental plant native to the Amazon Rainforest [38]. Like other Senna species, it is cultivated in humid and tropic regions of Africa, Asia, West Indices, Mexico, Australia, South America, the Caribbean Islands, Polynesia, Hawaii, Melanesia, and different parts of India [39]. In Philippines, Thailand, and Indonesia, this shrub is widely dispersed and is cultivated for medicinal purposes [40].

3.3. Ethnobotanical Uses. In Ayurvedic, Sinhala, Chinese, and African traditional medicine, various parts of S. alata have shown diverse therapeutic activities in diseases control. In northern Nigeria, the stem, leaf, and root decoctions is used in treatment of wound, skin and respiratory tract infection, burns, diarrhoea, and constipation [41, 42]. Also, in the south-western regions, leaf decoction serves as an antidote to body and abdominal pain, stress, and toothache [43]. It also cures dermal infections, convulsion, and as purgative [44, 45]. In Egypt, the leaf decoction was observed to be used as a bowel stimulant to stimulate peristaltic shrinkages and decrease water absorption from the colon in attempt to prevent constipation [46]. In Cameroon, the stem, bark, and leaves were reported to be used as a remedy for gastroenteritis, hepatitis, ringworm, and dermal infections [47].

In India, Philippines, and China, S. alata's stem, bark, and leaf decoction were found to be effective in treating haemorrhoids, inguinal hernia, syphilis, intestinal parasitosis, and diabetes [48]. Currently, seeds and roots are used in regulating uterus disorder and worms [18, 49]. The leaves and flowers are typically used as antifungal agents and laxatives [35]. In China, the seeds are used in treating asthma and to improve visibility [50]. In Guatemela, Brazil, and Guinea, the whole plant is currently used in the treatment of flu and malaria [36]. The flower and leaves decoction was observed to be used in treatment of ringworms, scabies, blotch, eczema, scabies, and tine infections [11, 51]. The leaves are currently used in Sierra Leone to relieve abortion pain and facilitate baby delivery [51]. In west and east Africa, bark decoction is spread on cuts during tribal mark incision and tattoo making [52]. In China, India, Benin republic, Ghana, Nigeria, and Togo, the whole plant is used as a curative for Diabetes mellitus [34, 53]. Fresh leaves of the plant are used in the treatment of skin rashes, mycosis, and dermatitis [54]. From the traditional uses, the frequent use of S. alata leaves is more than that of roots, flowers, roots, and stem-bark owing to more active metabolites reported in the leaves. Besides its pharmacological activities, it is processed into capsule, pellet, and tea in Nigeria for preventing diseases and maintaining sound health.

3.4. Pharmacological Activities. Medicinal plants belonging to the Fabaceae family have extensively been investigated for their pharmacological activities. Plants synthesised array of secondary metabolites which contribute to its therapeutic activities (Figure 1). Therapeutic appraisal of *S. alata*

Figure 1: Bioactive compounds with therapeutic potencies in S. alata.

authenticates the ethnobiological claims and establishes the pharmacological activities (Table 1). There are many published articles connected to diverse therapeutic activities of *S. alata* which are mainly related to antibacterial, antidiabetic, antilipogenic, antifungal, antioxidant, dermatophytic, antihyperlipidemic, and anthelmintic activities among others. Few studies have also reported its antimalarial activities.

3.4.1. Antibacterial Activities. The antibacterial potentials of medicinal herbs are appraised using zone of inhibition (ZOI) or minimum inhibitory concentration (MIC). The *in vivo* antibacterial potential of *S. alata* was assessed against methicillin-resistant *S. aureus* (MRSA), extended spectrum beta-lactamase, and carbapenemase-resistant. Enterobacteriaceae was isolated from infectious patients,

via Mueller-Hinton broth via the microdilution technique. The extract showed significant activities at 512 mg/ml due to the flavonoids, quinones, tannins, sterols, alkaloids, and saponins analysed [75]. S. alata leaves collected from an Indian aboriginal tribe, displayed significant ZOI of 21 to 27 mm against clinical isolates of multidrug-resistant (MDR) bacteria, obtained from infected patients, when subjected to ASTs via Kirbye Bauer's disc diffusion assay (KBDD) [58].

Ciprofloxacin (30 µg/disc) and vacuum liquid chromatographic (VLC) fractions of *S. alata* methanolic extract were assessed against *Staphylococcus aureus*, *Bacillus subtilis*, *Bacillus cereus* (Gram positive), *Shigella boydii*, *Shigella dysenteriae*, *Pseudomonas aureus*, *Vibrio mimicus*, *Salmonella paratyphi*, *Vibrio parahaemolyticus*, and *Salmonella typhi*, (Gram negative) using the disc diffusion assay. The

TABLE 1: Pharmacological activities of S. alata.

	Parts used	Country	Ethnomedicinal use	Solvent used	Pharmacological activity	Model used	Phytochemicals	References
(1)	Leaves	Nigeria	Treatment of diarrhoea, upper respiratory tract infection, and to hasten labour	Aqueous	Abortifacient	Pregnant rats	Saponins, flavonoids, cardiac glycosides, cardenolides, dienolides, phenolics and alkaloids	[55]
(2)	Leaves	India	To manage diabetes	Ethanolic	Hepato-renal protective effects	Male albino Wistar rats		[56]
(3)	Leaves	India	Treatment of allergy/asthma	Hydromethanolic	Antiallergic	Lipoxygenase (LOX) enzyme	Rhein and kaempferol	[7]
(4)	Leaves	Thailand	To manage diabetes and weight	Aqueous	Antilipogenic	High-fat diet- induced obese mice		[57]
(5)	Leaves	India	Treatment of bacterial infections	Methanol	Antibacterial	Pathogenic bacterial strains		[58]
(6)	Leaves	Cameroon	Treatment of gonorrhoea, gastrointestinal and skin diseases	Methanol	Antibacterial	Multidrug- resistant (MDR) Vibrio cholerae and Shigella flexneri	Kaempferol, luteolin and aloe-emodin	[59]
(7)	Leaves	India	To manage diabetes	Methanol	Antidiabetic	α-Glucosidase enzyme	Kaempferol 3- O-gentiobioside	[31]
(8)	Leaves	Thailand	Treatment of skin infections	Anthraquinone	Antifungal	STZ-induced diabetic rats	Anthraquinone	[60]
(9)	Leaves	Nigeria	To wash the uterus	Hexane	Anti-implantation, antigonadotropic, antiprogesteronic	Ovariectomized female rats	Alkaloids	[61]
(10)	Flower	Nigeria	Treatment of urinary tract infections and gonorrhoea	Methanol	Antibacterial	Bacterial strains	Steroids, anthraquinone glycosides, volatile oils and tannin	[62]
(11)	Flower	India	To treat scabies and ringworm	Aqueous	Antifungal	Aflatoxin producing and human pathogenic fungi		[63]
(12)	Leaves and barks	Malaysia	To treat superficial fungal infections	Ethanol and water	Antimicrobial	Bacterial and fungal strains		[64]
(13)	Leaves	Cameroon		Ethanol	Antioxidant and anti-inflammatory	White blood cells		[30]
(14)	Leaves	India	Used as purgative expectorant, astringent, vermicide	Ethanol	Anticancer	Male Wistar rats	Alkaloid, flavonoids, saponins, tannins glycosides	[65]
(15)	Leaves	Indonesia	malaria; antioxidant and antibacterial	Ethanol	Antiviral	DENV-2 and Huh 7it cells	Flavonoid	[66]
(16)	Petals	India	As an immune stimulant	Petroleum ether	Immunomodulatory	<i>Garra rufa</i> fish	Cardiac glycosides, phenols, anthraquinone, alkaloids	[67]

TABLE 1: Continued.

	Parts used	Country	Ethnomedicinal use	Solvent used	Pharmacological activity	Model used	Phytochemicals	References
(17)	Root	Nigeria	As an abortifacient in women, for the termination of	Ethanol	Uterine smooth muscle	Male and female Albino mice	Alkaloids	[68]
(18)	Leaves	Burkina Faso	early pregnancy Treatment of asthma-induced bronchospasm	Aqueous and ethanolic	Bronchorelaxant, genotoxic, and antigenotoxic	Male and female Wistar rats		[69]
(19)	Leaves	Egypt	Treatment of skin tumour	Methanol	Antitumor	Human cancer cell lines (HepG2, MDA- MB-231, and Caco2)	Palmitic, linolenic, linoleic, stearic acid	[70]
(20)	Leaves	Thailand	Laxative	Methanol	Anti-inflammatory	Tert-butyl hydroperoxide- induced oxidative stress in HaCaT cells	Rhein	[71]
(21)	Leaves, flower and fruit	Nigeria	Laxative and treatment of microbial infections	Methanol and ethanol	Antifungal and antibacterial	Clinical bacterial and fungal isolates	Flavonoids	[72]
(22)	Leaves	Thailand	In regulating glucose level in the blood	Aqueous	Antilipogenic	Male ICR mice		[73]
(23)	Leaves	Cameroon	To cure fever	Aqueous and methanolic	Antiplasmodial	RPMI 1640 and albumax		[74]

fractions exhibited significant inhibitory activities against bacteria isolates at 100 µg/ml [76].

Bioactive compounds such as kaempferol, luteolin, and aloe emodin isolated from the methanolic leaves extract was assessed against MDR bacteria strains. Aloe emodin exhibited strong MIC activities (4 to 128 µg/ml). The marked activities could be linked to the anthraquinone and flavonoids compounds detected [59]. The antibacterial activity of chrysoeriol-7-O-(2"-O-beta-D-mannopyranosyl)-beta-D-allopyranoside, 3-O-gentiobioside, quercetin, naringenin, and rhamnetin-3-O-(2"-O-beta-D-mannopyranosyl)-beta-D-allopyranoside isolated from chloroform, ethanol, petroleum ether, methanol, and aqueous leaves extract of *S. alata*, significantly inhibited the growth of *B. subtilis*, *V. cholerae*, *Streptococcus* sp., *S. aureus*, and *E. coli*. The extracts displayed pronounced activities which are in close proximity to penicillin, chloramphenicol, fluconazole, and ciprofloxacin [77].

Anthraquinone and flavonoid glycosides detected in *S. alata* extracts significantly inhibited the growth of *E. coli* and *S. aureus* with ZOI between 9.7 and 14.8 mm [22, 23]. The crude extracts and isolates obtained from purified fractions of *S. alata* flower extract (anthraquinone glycosides, steroids, tannins, and volatile oils) were assessed on selected bacteria isolates. Strong inhibitory activities were displayed at MIC of 500 mg/ml against the clinical isolates of *S. faecalis*, *B. subtilis*, *S. aureus*, *Pseudomonas putida*, and *M. Luteus*. At 500 mg/ml, ZOI of 10 to 25 mm was observed, with close proximity to the inhibitory activities displayed by streptomycin, penicillin, and methicillin [41]. The synergic effect of *Eugenia uniflora*

and *S. alata* leaves was examined on clinical isolates of *B. subtilis* and *S. aureus*. The dried leaves were processed into a local antiseptic herbal soap and the antibacterial potential was assessed via hole-in-plate agar diffusion assay. The herbal soap significantly inhibited the growth of the tested organisms [78].

3.4.2. Antioxidant Activities. The antioxidant or scavenging activities is important in appraisal of therapeutic potential of medicinal herbs. Plants played significant role in protecting cell against oxidative stress caused by active free radicals [79]. The free radical scavenging potentials of plants are appraised by antibiotic sensitivity tests (ASTs); 2,2-diphenyl-1-picrylhydrazyl (DPPH); ferric-reducing antioxidant power (FRAP); 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid) (ABTS); hydroxyl radical scavenging activity (HRSA), and metal ion chelating activity (MICA). The methanolic and n-hexane leaves extracts of S. alata were explored for their scavenging effects via DPPH assay. The methanolic extract displayed pronounced scavenging activities when compared to n-hexane extract. The methanolic extract was fractionated, leading to isolation of three major bioactive compounds (kaempferol, butylated hydroxytoluene, and emodin). The kaempferol fraction displayed pronounced scavenging activity than the butylated hydroxytoluene and emodin fractions [80]. Several bioactive compounds such as ascorbic acid, flavonoids, tocopherol, anthraquinone, and carotene assessed contributed to scavenging activities displayed [77].

Different parts of *S. alata*, significantly inhibited the action of free radicals causing oxidative stress [82]. The *in vitro* DPPH assay of ethyl acetate-DCM, methanol-DCM, and oil fractions of *S. alata* leaves was assessed using L-ascorbic acid as control. The radical scavenging activities measured at 517 nm significantly inhibited the free radicals at 500 g/ml. The concentration at 50% inhibition (IC $_{50}$) of the fractions revealed activities of methanol-DCM (65.03%), ethyl acetate-DCM (62.33%), and oil fraction (64.10%). The strong scavenging activities could be due to polyphenol and flavonoid analysed [76].

3.4.3. Antifungal Activities. Several bioactive compounds isolated from *S. alata* exhibit strong *in vitro* and *in vivo* antifungal activities. The antifungal activities of cannabinoid alkaloid (4-butylamine 10-methyl-6-hydroxy cannabinoid dronabinol), 1,8-cineole, caryophyllene, limonene, α -selinene, β -caryophyllene, germacrene D, hexadecanoic acid methyl ester, hexadecanoic acid, (6Z)-7,11-dimethyl-3-methylidenedodeca-1,6,10-triene, octadecanoic acid methyl ester, cinnamic acid, 3,7-dimethylocta-1,6-diene, pyrazol-5-ol, flavonol and gallic acid, methaqualone, and isoquinoline have been explored [51,83–85].

Volatile oils extracted from *S. alata* flowers were assessed against standard strains and clinical isolates of *Candida* and *Aspergillus* species. The oils significantly inhibited the growth of the tested microbes [86]. The methanolic extract and purified n-hexane and ethanolic fractions of *S. alata* flower displayed strong inhibitory activities against *A. Niger, C. utilis, G. candidum, A. brevipes,* and *Penicillium* species with an MIC of 0.312 to 5 mg/ml. However, at different concentrations, the purified fractions exhibited prominent inhibitory activities than the methanolic extracts. Likewise, mycelia growth was significantly inhibited by the purified fractions, with a total suppression of sporulation for 96 h at 2 mg/ml, compared to less sporulation after 48 h for methanolic extracts [87].

The aqueous and ethanolic leaf and bark extracts of *S. alata* was assessed via disc diffusion assay against *M. canis*, *C. albicans*, and *A. fumigatus*. The bark extracts displayed prominent concentration-dependent susceptibility against *C. albicans*. On the contrary, the aqueous extract showed a strong ZOI of 12 and 16 mm, while ethanolic extracts exhibited 10 and 14 mm. However, tioconazole displayed strong inhibitory activities of 18 mm which are significantly higher than the extracts at equivalent concentration [64].

 $S.\ alata$ flowers and leaves collected from Ogbomoso, Southwest Nigeria, were examined in an attempt to justify the indigenous claims of its antifungal efficacy. In vitro antifungal activities of ethanolic and methanolic extracts were investigated using the disc diffusion approach. Ethanolic extracts showed pronounced inhibitory activities when compared to methanolic extracts. The IC₅₀ value of the ethanolic extract was two folds higher than the methanolic extracts against the fungi isolates. The inhibitory activities displayed could be due to methaqualone, cinnamic acid, isoquinoline, and toluidine detected [71].

3.4.4. Dermatophytic Activities. Currently, the leaves, flowers, and bark of *S. alata* are used for treating various kinds of skin infections and diseases. In Thailand, the plant was mentioned as one of the 54 medicinal plants used for treating scabies, shingles, urticarial, itching, pityriasis versicolor, and ringworm [88]. The dermatophytic activities displayed by *S. alata* are linked to the bioactive compounds such as anthranols, anthrones, flavonoids, phenols, tannins, and anthracene derivatives [89]. Leaves decoction displayed strong inhibitory activities against *S. pyogenes, S. aureus, K. pneurnoniae, E. coli, S. rnarcescens, P. cepacia*, and *P. aeruginosa* [90, 91].

Cannabinoid alkaloid (4-butylamine 10-methyl-6-hydroxy cannabinoid dronabinol) and apigenin isolated from *S. alata* seeds were incorporated in a local antiseptic soap. The soap significantly hindered the spread of ringworms, eczemas, carbuncles, boils, infantile impetigo, and breast abscess [84]. The leaf, stem-bark, flower exudates, and ethanolic leaf extract examined against clinical isolates of *T. jirrucosum, M. canis, T. mentagrophyte, E. jlorrcosum, B. dermatitidis, A. flavus*, and *C. albicans* displayed strong inhibitory activities against the causative organisms [18,92–94].

3.4.5. Antilipogenic, Antidiabetic, and Antihyperlipidemic Activities. In Africa and Asia, S. alata leaves and flowers are currently used to regulate lipid absorption, obesity, and fat levels in blood serum. Aqueous leaf extract considerably reduced blood glucose levels, serum cholesterol, triglyceride, hepatic triglyceride, serum leptin, and insulin levels in Wistar mice [57].

Diabetes is a threat to man leading to stern socio-economic and high rate of mortality [95]. Herbal decoction of S. alata is commonly used in Caribbean [96], Africa [34], and India [97] for the treatment of diabetes. The marked α -glucosidase inhibitory effect and antidiabetic activities displayed by S. alata are linked to bioactive compounds such as astragalin and kaempferol-3-O-gentiobioside [31, 98]. Traditionally, the leaves decoction is used in regulation of sugar level in serum [99] and enhances the defence of hepato and renal organs to oxidative stress [56]. It also reduces fasting blood glucose and improves metabolic state via activation of PKB/Akt which helps regulate glucose uptake in the liver and muscles [100].

The antidiabetic potency of acarbose, n-butanol, and ethyl acetate fractions of methanolic *S. alata* leaf extract was assessed via inhibitory carbohydrate digestion mechanism. The antidiabetic drug (acarbose) significantly inhibits α -glucosidase with inhibitory concentration (IC50, $107.31 \pm 12.31 \,\mu\text{g/ml}$). n-Butanol and ethyl acetate fractions displayed inhibitory activities of IC50, $2.5.80 \pm 2.01 \,\mu\text{g/ml}$ and IC50, $2.95 \pm 0.47 \,\mu\text{g/ml}$, respectively, due to kaempferol 3-O-gentiobioside analysed by HPLC and Combiflash chromatography [31, 83]. α -Amylase and α -glucosidase inhibitory potential of acetone, hexane, and ethyl acetate leaf extracts of *S. alata* was assessed via Lineweaver–Burk plot. Acetone extract significantly inhibited α -amylase (IC50 = 6.41 mg/mL), while hexane displayed strong inhibition against α -glucosidase (IC50 = 0.85 mg/mL). n-Hexane

substantially reduces (p < 0.05) postprandial blood glucose in sucrose-induced rats after 2 h of treatment [101].

The reduction in the oxidative stress in the aorta and heart of streptozotoc, hyperglycemic rats was investigated by DPPH free radical scavenging and antioxidant catalase assays. The aqueous leaf extract significantly reduced lipid peroxidation (MDA levels), blood glucose; however, it increases in the antioxidant catalase and DPPH free radical scavenging activity was observed. The reversal of oxidative stress induced by cardiac dysfunction in hyperglycemic condition justified the ethnobiological claim of the plant [102]. In a similar study, the *S. alata* leaf extract significantly reduces oxidative stress causing diabetes in treated mice. Prominent activities were observed in the kidney and liver. The activity resulted in critical changes in the urea, protein, creatinine, and uric acid level in the blood serum [56].

3.4.6. Antimalarial Activities. Malaria is a global threat contributing to serious sociocultural and health issues in humid and tropical regions [103, 104]. Chemotherapy reported the antimalarial activities of S. alata could be linked to quinones, alkaloids, and terpenes [105, 106]. Quinones isolated from S. alata significantly displayed in vitro antiplasmodial activity, against Plasmodium falciparum via the microdilution test of Desjardin [107]. Terpenes isolated from S. alata leaves displayed pronounce in vitro antiplasmodial assays against P. falciparum in ethylene glycol-water fractions. Significant activity was observed at concentration below $1 \mu g/ml$ [108]. The appraisal of aqueous leaf extract justifies the ethnomedical applications of S. alata as remedy for malaria and fever. The leaf extract considerably inhibited 3D7 strain of the P. falciparum parasite in Wistar mice [109].

3.4.7. Anthelmintic Activity. Traditionally, S. alata leaf and flower decoctions are used in treatment of intestinal worm infestation and stomach disorder. The anthelmintic potency of alcoholic leaves extract of S. alata and T. angustifolia at 10 to 100 mg/ml were assessed in clinical isolates of Ascardia galli and Pheretima posthuma by observing time of paralysis and point of death of the worms. The leaves extract significantly inhibit the worms (test organisms) more than piperazine citrate (standard anthelmintic drug) [110].

3.4.8. Antiviral Activities. S. alata is an indispensable bactericidal and fungicidal natural therapy. However, the justification of antiviral activities is not properly documented. The antiviral efficacy of n-hexane, ethyl acetate, butanol, and aqueous leaf extracts was assessed on dengue virus (DENV) obtained from an infected pregnant woman in Indonesia via focus assay. The extracts significantly inhibited DENV-2 with IC₅₀ (<10 μ g/ml), CC₅₀ (645.8 μ g/ml) and SI (64.5 μ g/ml) [111].

4. Nutritional Constituents of S. alata

In appraisal of the therapeutic potentials of medicinal herbs, phytochemical, pharmacological, toxicological, and

nutritional details are indispensable. In appraisal of the nutritional constituents of S. alata leaf, several nutrients assessed include moisture content (9.53 ± 0.06) , carbohydrate (298.61 ± 0.40) , crude lipid (47.73 ± 0.01) , crude fibre (18.23 ± 0.13) , and ash (15.73 ± 0.03) . The nutritional contents analysed in flower revealed ash (7.00 ± 1.0) , crude protein (13.14 ± 0.02) , carbohydrate (57.04 ± 0.04) , moisture (6.16 ± 0.14) , and crude lipid (1.81 ± 0.09) . It is observed that storage capacity, easy absorption of food, antimicrobial and anaesthetic properties are connected to the nutritional composition of herbs [112]. Several minerals such as potassium, iron, manganese, calcium, zinc, copper, and chromium have been assessed in S. alata. These mineral constituents help in formulation of herbal drugs and dietary supplements [113-115].

5. Toxicological Studies of S. alata

Lately, medicinal plants and herbal products have come under spotlight owing to health hazards connected to indiscriminate handling of herbs and herbal products. Several reports were published relating to the use of herbal drugs. Despite the risks ascribed to most medicinal and herbal products, the number of herbal users still increases. This initiates the appraisal of herbs in an attempt to establish their safety [116]. The S. alata leaf extract was appraised by oral administration of Wistar rats (10 g/kg weight). The acute and subacute effects were assessed in every 24 h for 14 days at different dosages (250, 500, and 1000 mg/kg). After 24 h, no significant variation was observed in the liver, urea, bicarbonate, creatinine, and kidney of the tested organisms and control [117]. Aqueous flower extract at doses of 100, 400, and 800 mg/kg was orally administered to ten Wistar rats for 28 days. No significant difference on histological sections of the lung, liver, kidney, heart, and kidney was observed during the postmortem analysis of the mice [118].

Different doses (1,000, 2,000, and 3,000 mg/kg body weight) of ethanolic leaf extract do not display any behavioural, biochemical, haematological, and histopathological variations in the tested Swiss albino mice and control after 15 days of oral administration [119]. Hydroethanolic leaf extract increased the body weight, blood level, liver homogenates, and serum index after proper monitoring of the administered rats every 48 h for 26 days. Haematological parameters assessed displayed dose-dependent response, however, no significant variation was observed in liver histopathology [120]. Alkaloids isolated from aqueous S. alata leaf extract significantly reduces alkaline phosphate, uric acid, globulin, liver and kidney body weight ratio, urea, phosphate ions, bilirubin, potassium ions, serum concentrations of albumin, creatinine, gamma glutamyl transferase, alanine transaminases, and aspartate in the liver and kidney of pregnant rats after periodic observation for 18 days. This led to variation in serum enzymes, excretory, and secretory functions of liver and kidney of the test organisms [61].

Documented toxicological appraisals of *S. alata* present positive information regarding its safety; however, there is need to authenticate these reports through *in vitro* and *in vivo* assessments on clinical isolate. Fresh and dried plant

parts, low-dosage concentrations, test organisms of different age groups and effect on other vital organs such as brain, intestine, and bladder should be considered during toxicological appraisals of medicinal plant or products. Ethnobiological appraisals described that ulcer patients, pregnant women, and children under the age of two should avoid *S. alata* decoction. Also, studies should be focused on appraisals of recommended dosage, safest mode of administration, and reaction mechanisms of the metabolites in living organisms.

6. Limitation, Conclusion, and Future Prospects

Based on this review, further studies on S. alata have become evident. Most studies focused on lipid-soluble and volatile components; however, appraisals on metabolites prepared in hydrophilic medium are deficient. Most phytochemical studies of S. alata focused on leaves, stems, and flowers, though study on seeds and roots are lacking. Many therapeutic appraisals have been investigated in cell and animal experiments; still, there remain important tasks. These include secondary metabolites dictating the pharmacological activities are not well stated; relatively, few studies analytically appraised the mechanisms of action of secondary metabolites; systemic toxicological studies remain ambiguous. The phytochemical appraisals revealed there is a variation in some metabolites investigated due to geographical and climatic factors. This gives room for qualitative approach to authenticate the metabolites in this plant. The plant secondary metabolites contributed to the pronounced wound healing and antibacterial and antifungal activities [92, 121]. Industrially, the plant is a key ingredient in detergents (foaming and surface active agents), molluscicides and pesticides. Summarily, comprehensive ethnobiological and pharmacological activities of S. alata, limitations, and future prospects were explored which could present novel research directions.

Abbreviations

ZOI: Zone of inhibition

MIC: Minimum inhibitory concentration

MDR: Multidrug resistant

ASTs: Antibiotic sensitivity tests
DPPH: 2,2-Diphenyl-1-picrylhydrazyl
FRAP: Ferric-reducing antioxidant power
ABTS: Chemiluminescence, 2,2'-azino-bis(3-

ethylbenzothiazoline-6-sulphonic acid)

HRSA: Hydroxyl radical scavenging activity

MICA: Metal ion chelating activity.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

The authors are indebted to the Department of Physical Sciences, Faculty of Pure and Applied Sciences, Landmark University, Omu-Aran, Nigeria.

References

- [1] O. S. Oladeji, K. A. Odelade, and J. K. Oloke, "Phytochemical screening and antimicrobial investigation of *Moringa oleifera* leaf extracts," *African Journal of Science, Technology, Innovation and Development*, vol. 12, no. 1, pp. 79–84, 2019.
- [2] M. Ekor, "The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety," *Frontier of Pharmacology*, vol. 4, pp. 1–10, 2013.
- [3] WHO (World Health Organization), Global Report on Diabetes, WHO, Geneva, Switzerland, 2016.
- [4] R. Verpoorte, "Forward," in Evidence-Based Validation of Herbal Medicine, P. K. Mukherjee, Ed., pp. 13-14, Elsevier, Amsterdam, Netherlands, 2015.
- [5] S. Svahn, "Analysis of secondary metabolites from Aspergillus fumigatus and Penicillium nalgiovense," Digital Comprehensive Summaries of Uppsala Dissertations from the Faculty of Pharmacy, p. 195, Uppsala University, Uppsala, Sweden, 2015.
- [6] M. J. Balunas and A. D. Kinghorn, "Drug discovery from medicinal plants," *Life Sciences*, vol. 78, no. 5, pp. 431–441, 2005
- [7] B. Singh, J. R. Nadkarni, R. A. Vishwakarma, S. B. Bharate, M. Nivsarkar, and S. Anandjiwala, "The hydroalcoholic extract of *Cassia alata* (Linn.) leaves and its major compound rhein exhibits antiallergic activity via mast cell stabilization and lipoxygenase inhibition," *Journal of Ethnopharmacology*, vol. 141, no. 1, pp. 469–473, 2012.
- [8] C. Veeresham, "Natural products derived from plants as a source of drugs," *Journal of Advanced Pharmaceutical Technological Research*, vol. 3, no. 4, pp. 200-201, 2012.
- [9] A. Kumar, R. Shukla, P. Singh, C. S. Prasad, and N. K. Prasad, "Assessment of Thymus vulgaris L. essential oil as a safe botanical preservative against post harvest fungal infestation of food commodities," *Innovative Food Science & Emerging Technologies*, vol. 9, no. 4, pp. 575–580, 2008.
- [10] A. Liu, L. Xu, Z. Zou et al., "Studies on chemical constituents from leaves of Cassia alata," China Journal of Chinese Matters on Medicine, vol. 34, pp. 861–863, 2009.
- [11] C. P. Khare, *Indian Medicinal Plants: An Illustrated dictionary*, Springer Press, London, UK, 2007.
- [12] C. V. Chang, A. C. Felício, J. E. D. P. Reis, M. D. O. Guerra, and V. M. Peters, "Fetal toxicity of *Solanum lycocarpum* (Solanaceae) in rats," *Journal of Ethnopharmacology*, vol. 81, no. 2, pp. 265–269, 2002.
- [13] M. Ackland, S. van de Waarsenburg, and R. Jones, "Synergistic antiproliferative action of the flavonols quercetin and kaempferol in cultured human cancer cell lines," *In Vivo*, vol. 19, pp. 69–76, 2004.
- [14] E. J. Choi and W. S. Ahn, "Kaempferol induced the apoptosis via cell cycle arrest in human breast cancer MDA-MB-453 cells," *Nutrition Research and Practice*, vol. 2, no. 4, pp. 322–325, 2008.
- [15] S. Bandyopadhyay, J. R. Romero, and N. Chattopadhyay, "Kaempferol and quercetin stimulate granulocyte-macrophage colony-stimulating factor secretion in human prostate cancer cells," *Molecular and Cellular Endocrinology*, vol. 287, no. 1-2, pp. 57–64, 2008.
- [16] V. E. Fernand, J. N. Losso, R. E. Truax et al., "Rhein inhibits angiogenesis and the viability of hormone-dependent and -independent cancer cells under normoxic or hypoxic conditions in vitro," Chemico-Biological Interactions, vol. 192, no. 3, pp. 220–232, 2011.

- [17] G. Aviello, I. Rowland, C. I. Gill et al., "Anti-proliferative effect of rhein, an anthraquinone isolated from Cassia species, on Caco-2 human adenocarcinoma cells," *Journal of Cellular and Molecular Medicine*, vol. 14, pp. 2006–2014, 2009
- [18] A. A. Makinde, J. O. Igoli, L. TA'Ama et al., "Antimicrobial activity of Cassia alata," African Journal of Biotechnology, vol. 6, no. 13, pp. 1509-1510, 2007.
- [19] P. A. Tarkang, G. A. Agbor, T. D. Armelle et al., "Acute and chronic toxicity studies of the aqueous and ethanol leaf extracts of *Carica papaya* Linn in Wistar rats," *Journal of Natural Product and Plant Resources*, vol. 2, no. 5, pp. 617–627, 2012.
- [20] K. Aftab, M. D. Ali, P. Aijaz et al., "Determination of different trace and essential element in lemon grass samples by X-ray flouresence spectroscopy technique," *International Food Research Journal*, vol. 18, pp. 265–270, 2011.
- [21] I. M. Villaseñor, A. P. Canlas, M. Pascua et al., "Bioactivity studies on *Cassia alata* Linn. leaf extracts," *Phytotherapy Research*, vol. 16, no. 1, pp. 93–96, 2002.
- [22] M. R. Khan, M. Kihara, and A. D. Omoloso, "Antimicrobial activity of *Cassia alata*," *Fitoterapia*, vol. 72, no. 5, pp. 561–564, 2001.
- [23] M. N. Somchit, I. Reezal, I. E. Nur, and A. R. Mutalib, "In vitro antimicrobial activity of ethanol and water extracts of *Cassia alata*," *Journal of Ethnopharmacology*, vol. 84, no. 1, pp. 1–4, 2003.
- [24] E. O. Donatus and U. N. Fred, "Cannabinoid Dronabinol alkaloid with antimicrobial activity from *Cassia alata Linn*," *Der Chemica Sinica*, vol. 2, no. 2, pp. 247–254, 2010.
- [25] S. Ranganathan and S. A. Balajee, "Anti-cryptococcus activity of combination of extracts of *Cassia alata* and *Ocimum sanctum*," *Mycoses*, vol. 43, no. 7-8, pp. 299–301, 2000.
- [26] R. B. Otto, S. Ameso, and B. Onegi, "Assessment of anti-bacterial activity of crude leaf and root extracts of *Cassia alata* against *Neisseria gonorrhea*," *African Health Science*, vol. 14, pp. 840–848, 2014.
- [27] S. Gaikwad, "Isolation and characterization of a substituted Anthraquinone: a bioactive compound from *Cassia auriculata* L," *Industrial Journal of Advanced Plant Research*, vol. 1, no. 5, pp. 8–11, 2014.
- [28] E. I. Olarte, A. A. Herrera, I. M. Villasenor et al., "Anti-oxidant activity of a new indole alkaloid from *Cassia alata L*," *Philippine Agricultural Scientist*, vol. 93, no. 3, pp. 250–254, 2010.
- [29] M. Manogaran and N. Sulochana, "Anti-inflammatory activity of Cassia aauriculata," Science Life, vol. 24, no. 2, pp. 65–67, 2004.
- [30] B. Sagnia, D. Fedeli, R. Casetti et al., "Antioxidant and antiinflammatory activities of extracts from *C. alata, E. indica, E. speciosa, C. papaya* and *P. fulva* medicinal plants collected in Cameroon," *PLoS One*, vol. 9, no. 8, Article ID e103999, 2014.
- [31] G. K. Varghese, L. V. Bose, and S. Habtemariam, "Antidiabetic components of Cassia alata leaves: identification through α-glucosidase inhibition studies," *Pharmaceutical Biology*, vol. 51, no. 3, pp. 345–349, 2013.
- [32] S. M. Midawa, B. D. Ali, B. Z. Mshelia et al., "Cutaneous wound healing activity of the ethanolic extracts of the leaf of Senna alata L (Caesalpiniaceae)," Journal of Biological Science and Bioconservation, vol. 2, 2010.
- [33] S. Kundu, S. Roy, and L. M. Lyndem, "Cassia alata L: potential role as anthelmintic agent against Hymenolepis diminuta," *Parasitology Research*, vol. 111, no. 3, pp. 1187–1192, 2012.

- [34] K. A. Abo, A. A. Fred-Jaiyesimi, and A. E. A. Jaiyesimi, "Ethnobotanical studies of medicinal plants used in the management of diabetes mellitus in South Western Nigeria," *Journal of Ethnopharmacology*, vol. 115, no. 1, pp. 67–71, 2008.
- [35] N. R. Farnsworth and N. Bunyapraphatsara, Thai Medicinal Plants. Recommended for Primary Health Care System, Medicinal Plant Information Center, Faculty of Pharmacy, Mahidol University, Salaya, Thailand, 1992.
- [36] T. Hennebelle, B. Weniger, H. Joseph, S. Sahpaz, and F. Bailleul, "Senna alata," *Fitoterapia*, vol. 80, no. 7, pp. 385–393, 2009.
- [37] V. R. Kumar, S. Kumar, S. Shashidhara et al., "Comparison of the antioxidant capacity of an important hepatoprotective plants," *International Journal of Pharmaceutical Science and Drug Research*, vol. 3, pp. 48–51, 2011.
- [38] O. S. Oladeji, F. E. Adelowo, D. T. Ayodele, and K. A. Odelade, "Phytochemistry and pharmacological activities of *Cymbopogon citratus*: a review," *Scientific African*, vol. 6, Article ID e00137, 2019.
- [39] I. A. Ross, Medicinal Plants of the World, Humana Press, Totowa, NJ, USA, 1999.
- [40] S. Palanichamy and S. Nagarajan, "Analgesic activity of *Cassia alata* leaf extract and kaempferol 3-o-sophoroside," *Journal of Ethnopharmacology*, vol. 29, no. 1, pp. 73–78, 1990.
- [41] O. Adedayo, W. A. Anderson, M. Moo-Young, V. Snieckus, P. A. Patil, and D. O. Kolawole, "Phytochemistry and antibacterial activity of Senna alata flower," *Pharmaceutical Biology*, vol. 39, no. 6, pp. 408–412, 2001.
- [42] A. M. El-Mahmood, J. H. Doughari, and F. J. Chanji, "In vitro antibacterial activities of crude extract of Nauclea latifolia and Daniella oliver," Scientific Research Essays, vol. 31, no. 3, pp. 102–105, 2003.
- [43] T. V. Benjamin and A. Lamikanra, "Investigation of Cassia alata, a plant used in Nigeria in the treatment of skin diseases," *Quarterly Journal of Crude Drug Research*, vol. 19, no. 2-3, pp. 93–96, 2008.
- [44] S. O. Oguntoye, J. A. Owoyale, and G. A. Olatunji, "Antifungal and antibacterial activities of an alcoholic extract of *S. alata* leaves," *Journal of Applied Science and Environmental Management*, vol. 9, no. 3, pp. 105–107, 2005.
- [45] E. O. Ogunti and A. A. Elujobi, "Laxative activity of *Cassia alata*," *Fitoterapia*, vol. 64, no. 5, pp. 437–439, 1993.
- [46] L. Leung, T. Riutta, J. Kotecha, and W. Rosser, "Chronic constipation: an evidence-based review," *The Journal of the American Board of Family Medicine*, vol. 24, no. 4, pp. 436–451, 2011.
- [47] R. A. Day and A. L. Underwood, *Quantitative Analysis*, Prentice-Hall Publication, London, UK, 5th edition, 1986.
- [48] J. O. Igoli, O. G. Ogaji, T. A. Tor-Anyiin et al., "Traditional medicinal practices among the Igede people of Nigeria (part II)," *African Journal of Traditional Complementary and Alternative Medicine*, vol. 2, no. 2, pp. 134–152, 2005.
- [49] V. Selvi, V. Isaivani, and S. Karpagam, "Studies on antimicrobial activities from flower extract of *Cassia alata Linn*," *International Journal of Current Science*, vol. 2, pp. 299–303, 2012.
- [50] K. N. Chauhan, M. B. Patel, H. R. Valera et al., "Hepatoprotective activity of flowers of *Cassia auriculata* R. against paracetamol induced liver injury," *Journal of Natural Remedies*, vol. 9, pp. 85–89, 2009.
- [51] S. O. Oladeji, F. E. Adelowo, and K. A. Odelade, "Mass spectroscopic and phytochemical screening of phenolic compounds in the leaf extract of Senna alata (L.) Roxb.

- (Fabales: Fabaceae)," *Brazilian Journal of Biological Sciences*, vol. 3, no. 5, pp. 209–219, 2016.
- [52] J. M. Dalziel, Useful Plants of West Tropical Africa, Crown Agents for Overseas Governments, London, UK, 1956.
- [53] W. Woradulayapinij, N. Soonthornchareonnon, and C. Wiwat, "In vitro HIV type 1 reverse transcriptase inhibitory activities of Thai medicinal plants and Canna indica L. rhizomes," *Journal of Ethnopharmacology*, vol. 101, no. 1–3, pp. 84–89, 2005.
- [54] K. K. Ajibesin, B. A. Ekpo, D. N. Bala, E. E. Essien, and S. A. Adesanya, "Ethnobotanical survey of Akwa ibom state of Nigeria," *Journal of Ethnopharmacology*, vol. 115, no. 3, pp. 387–408, 2008.
- [55] M. T. Yakubu, A. O. Adeshina, A. T. Oladiji et al., "Abortifacient potential of aqueous extract of *S. alata* leaves in rats," *Journal of Reproduction and Contraception*, vol. 21, no. 3, pp. 163–177, 2010.
- [56] M. Sugumar, D. Victor, A. Dossb et al., "Hepato-renal protective effects of hydroethanolic extract of *S. alata* on enzymatic and nonenzymatic antioxidant systems in streptozotocin induced diabetic rats," *Integrative Medicine Research*, vol. 5, pp. 276–283, 2016.
- [57] J. Naowaboot, N. Somparn, S. Saentaweesuk et al., "Umbelliferone improves an impaired glucose and lipid metabolism in high-fat diet/streptozotocin-induced type 2 diabetic rats," *Phytotherapy Research*, vol. 29, pp. 1388–1395, 2015.
- [58] S. S. Swain and R. N. Padhy, "In vitro antibacterial efficacy of plants used by an Indian aboriginal tribe against pathogenic bacteria isolated from clinical samples," *Journal of Taibah University Medical Sciences*, vol. 10, no. 4, pp. 379–390, 2015.
- [59] S. J. N. Tatsimo, J.-d.-D. Tamokou, V. T. Tsague et al., "Antibacterial-guided isolation of constituents from Senna alata leaves with a particular reference against multi-drugresistant Vibrio cholerae and Shigella flexneri," *International Journal of Biological and Chemical Sciences*, vol. 11, no. 1, pp. 46–53, 2017.
- [60] M. Wuthi-udomlert, P. Kupittayanant, and W. Gritsanapan, "*In vitro* evaluation of antifungal activity of anthraquinone derivatives of *S. alata*," *Journal of Health Research*, vol. 24, no. 3, pp. 117–122, 2010.
- [61] M. T. Yakubu and I. F. Musa, "Effects of post-coital administration of alkaloids from S. alata (Linn. Roxb) leaves on some fetal and maternal outcomes of pregnant rats," Journal of Reproduction and Infertility, vol. 13, no. 4, pp. 211–217, 2012.
- [62] M. N. Abubacker, R. Ramanathan, and T. S. Kumar, "In vitro antifungal activity of Cassia alata Linn flower extract," Natural Products Radiance, vol. 7, no. 1, pp. 6–9, 2008.
- [63] N. M. Kudatarkar and Y. K. Nayak, "Pharmacological screening of Cassia alata leaves on colorectal cancer," Colorectal Cancer, vol. 4, no. 1, p. 2, 2018.
- [64] M. N. Somchit and A. R. Mutalib, "Tropical medicine," Plants, vol. 2, no. 2, pp. 179–182, 2003.
- [65] A. Marissa, H. Muhammad, D. Franciscus et al., "Antiviral effect of sub fraction *Cassia alata* leaves extract to dengue virus serotype-2 strain new guinea C in human cell line huh-7 it-1," *IOP Conference Series: Earth and Environmental Science*, vol. 101, Article ID 012004, 2017.
- [66] R. Jayasree, R. Prathiba, and S. Sangavi, "Immunomodulatory effect of Cassia alata petals in Garra rufa (doctor fish)," Journal of Chemical and Pharmaceutical Sciences, vol. 9, no. 1, pp. 215–218, 2016.

- [67] T. A. Adedoyin, I. O. Joshua, O. E. Ofem et al., "Effects of Cassia alata root extract on smooth muscle activity," British Journal of Pharmaceutical Research, vol. 5, no. 6, pp. 406–418, 2015
- [68] M. Ouédraogo, F. Da, A. Fabré et al., "Evaluation of the bronchorelaxant, genotoxic, and antigenotoxic effects of Cassia alata L," Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 162651, 11 pages, 2013.
- [69] M. M. Mohammed, S. S. El-Souda, S. M. El-Hallouty et al., "Antiviral and cytotoxic activities of anthraquinones isolated from *Cassia roxburghii* Linn. leaves," *Herba Polonica*, vol. 59, no. 4, pp. 33–44, 2013.
- [70] K. Wadkhien, C. Chinpaisal, M. Satiraphan et al., "Anti-inflammatory effects of rhein and crude extracts from *Cassia* alata L. in HaCaT cells," *Science, Engineering and Health Studies*, vol. 12, no. 1, pp. 19–32, 2018.
- [71] F. Adelowo and O. Oladeji, "An overview of the phytochemical analysis of bioactive compounds in *S. alata*," *American Chemical and Biochemical Engineering*, vol. 2, no. 1, pp. 7–14, 2017.
- [72] J. Naowaboot and P. Piyabhan, "S. alata leaf extract restores insulin sensitivity in high-fat diet-induced obese mice," Clinical Phytoscience, vol. 2, p. 18, 2016.
- [73] W. P. Josué, N. A. Nadia, K. M. Claire et al., "In Vitro sensitivity of *Plasmodium falciparum* field isolates to methanolic and aqueous extracts of *Cassia alata* (Fabaceae)," *Alternative Integrative Medicine*, vol. 3, p. 2, 2014.
- [74] G. E. Trease and W. C. Evans, "Phenols and phenolic gly-cosides," in *Pharmacognosy*, G. E. Trease and W. C. Evans, Eds., pp. 832-833, Biliere Tindall, London, UK, 1996.
- [75] P. Wikaningtyas and E. Y. Sukandar, "The antibacterial activity of selected plants towards resistant bacteria isolated from clinical specimens," *Asian Pacific Journal of Tropical Biomedicine*, vol. 6, no. 1, pp. 16–19, 2016.
- [76] B. K. Saha, M. N. Bhuiyan, K. Mazumder et al., "Hypogly-cemic activity of *Lagerstroemia speciose* L. extract on streptozotocin-induced diabetic rat: underlying mechanism of action," *Bangladesh Journal of Pharmacology*, vol. 4, no. 2, pp. 79–83, 2009.
- [77] S. Chatterjee and S. Dutta, "An Overview on the ethnophytopathological studies of *Cassia alata*-an important medicinal plant and the effect of VAM on its growth and productivity," *International Journal of Research on Botany*, vol. 2, no. 4, pp. 13–19, 2012.
- [78] O. A. Oyedele, C. N. Ezekiel, M. Sulyok et al., "Mycotoxin risk assessment for consumers of groundnut in domestic markets in Nigeria," *International Journal of Food Micro*biology, vol. 251, pp. 24–32, 2017.
- [79] Y. Z. Zhu, S. H. Huang, B. K. Tan et al., "Antioxidants in Chinese herbal medicines: a biochemical perspective," *Natural Products Representation*, vol. 21, pp. 478–489, 2004.
- [80] P. Panichayupakaranant and S. Kaewsuwan, "Bioassay-guided isolation of the antioxidant constituent from Cassia alata L. leaves," Songklanakarin Journal of Science and Technology, vol. 26, no. 1, pp. 103–107, 2004.
- [81] S. Chatterjee, S. Chatterhee, K. Dey et al., "Study of anti-oxidant activity and immune stimulating potency of the ethnomedicinal plant, *Cassia alata* (L.) Roxb," *Medical Aromatic Plants*, vol. 2, no. 4, pp. 1–6, 2013.
- [82] B. Sarkar, S. Khodre, P. Patel et al., "HPLC analysis and antioxidant potential of plant extract of *Cassia alata*," *Asian Journal of Pharmaceutical Science and Technology*, vol. 4, no. 1, pp. 4–7, 2014.

- [83] O. T. Tcheghebe, V. R. Tala, F. N. Tatong et al., "Ethnobotanical uses, phytochemical and pharmacological profiles, and toxicity of *Cassia alata L.* an overview," *Landmark Research Journal of Medicine and Medical Science*, vol. 4, no. 2, pp. 16–24, 2017.
- [84] D. E. Okwu and F. U. Nnamdi, "Cannabinoid dronabinol alkaloid with antimicrobial activity from Cassia alata Linn," Der Chemica Sinica, vol. 2, no. 2, pp. 247–254, 2011.
- [85] I. A. Ogunwande, G. Flamini, P. L. Cioni et al., "Aromatic plants growing in Nigeria: essential oil constituents of Cassia alata (Linn.) roxb. And Helianthus annuus L.," Records of Natural Products, vol. 4, no. 4, pp. 211–217, 2010.
- [86] E. E. Essien, T. M. Walker, I. A. Ogunwande et al., "Volatile constituents, antimicrobial and cytotoxicity potentials of three Senna species from Nigeria," *Journal of Essential Oil Bearing Plants*, vol. 14, no. 6, pp. 722–730, 2011.
- [87] O. Adedayo, W. A. Anderson, and M. Moo-Young, "Antifungal properties of some components of *S. alata* flower," *Pharmaceutical Biology*, vol. 37, no. 5, pp. 369–374, 1999.
- [88] O. Neamsuvan, "A survey of herbal formulas for skin diseases from Thailand's three southern border provinces," *Journal of Herbal Medicine*, vol. 5, no. 4, pp. 190–198, 2015.
- [89] E. Ramstad, Modern Pharmacognosy, McGraw-Hill, New York, NY, USA, 1959.
- [90] S. H. Brown, S. alata, The Institute of Food and Agricultural Sciences, University of Florida, Gainesville, FL, USA, 2011.
- [91] T. V. Benjamin and A. Lamikanra, "Investigation of *Cassia alata*, a plant used in Nigeria in the treatment of skin diseases," *Quarterly Journal of Crude Drug Research*, vol. 19, no. 2-3, pp. 93–96, 1981.
- [92] W. F. Sule, I. O. Okonko, T. A. Joseph et al., "In vitro antifungal activity of *S. alata* Linn. crude leaf extract," *Research Journal of Biological Science*, vol. 5, no. 3, pp. 275–284, 2010.
- [93] N. Ali-Emmanuel, M. Moudachirou, J. A. Akakpo et al., "Treatment of bovine dermatophilosis with Senna alata, Lantana camara and Mitracarpus scaber leaf extracts," Journal of Ethnopharmacology, vol. 86, pp. 167–217, 2003.
- [94] W. F. Sule, I. O. Okonkwo, S. Omo-Ogun et al., "Phytochemical properties and *in vitro* antifungal activity of *Senna alata* Linn. crude stem bark extract," *Journal of Medicinal Plants Research*, vol. 5, no. 2, pp. 176–183, 2011.
- [95] U. F. Ezuruike and J. M. Prieto, "The use of plants in the traditional management of diabetes in Nigeria: pharmacological and toxicological considerations," *Journal of Ethnopharmacology*, vol. 155, no. 2, pp. 857–924, 2014.
- [96] L. M. Girón, V. Freire, A. Alonzo et al., "Ethnobotanical survey of the medicinal flora used by the caribs of guatemala," *Journal of Ethnopharmacology*, vol. 34, no. 2-3, pp. 173–187, 1991.
- [97] M. H. Khan and P. S. Yadava, "Antidiabetic plants used in Thoubal district Nanipur. Northeast India," *Indian Journal of Traditional Knowledge*, vol. 9, pp. 510–514, 2010.
- [98] T. A. Adelusi, G. Oboh, J. A. Ayodele et al., "Avocado pear fruits and leaves aqueous extracts inhibit alpha-amylase, alpha-glucosidase and SNP induced lipid peroxidation—an insight into mechanisms involve in the management of type 2 diabetes," *International Journal of Applied Natural Science*, vol. 3, no. 5, pp. 21–34, 2014.
- [99] S. Palanichamy, S. Nagarajan, and M. Devasagayam, "Effect of Cassia alata leaf extract on hyperglycemic rats," *Journal of Ethnopharmacology*, vol. 22, pp. 81–90, 1988.
- [100] C. R. Lima, C. F. Vasconcelos, J. H. Costa-Silva et al., "Anti-diabetic activity of extract from *Persea americana* mill leaf via the activation of protein kinase B (PKB/Akt) in

- streptozotocin-induced diabetic rats," *Journal of Ethno-pharmacology*, vol. 141, pp. 517–525, 2012.
- [101] M. I. Kazeem, G. Azeez, and O. Ashafa, "Effect of S. alata (L) Roxb (Fabaceae) leaf extracts on alpha-amylase, alpha-glucosidase and postprandial hyperglycemia in rats," Tropical Journal of Pharmaceutical Research, vol. 14, no. 10, pp. 1843–1848, 2015.
- [102] I. Reezal, M. N. Somchif, and R. M. Abdul, "In vitro antifungal properties of Cassia alata," in Proceedings of the Regional Symposium on Environment and Natural Resources, Hotel Renaissance Kuala Lumpur, Kuala Lumpur, Malaysia, pp. 654–659, April 2002.
- [103] R. Singh, N. Kaur, L. Kishore et al., "Management of diabetic complications: a chemical constituents based approach," *Journal of Ethnopharmacology*, vol. 150, pp. 51–70, 2013.
- [104] WHO, World Malaria Report, World Health Organization, Geneva, Switzerland, 2014.
- [105] S. Federici, R. Quaratino, D. Papale et al., Sistema Informatico delle Tavole Alsometriche d'Italia, DiSAFRi, Università Degli Studi Della Tuscia, Viterbo, Italy, 2001, http://gaia. agraria.unitus.it.
- [106] R. E. Fowler, P. F. Billingsley, M. Pudney et al., "Inhibitory action of the antimalarial compound atovaquone (566C80) against *Plasmodium berghei* ANKA in mosquito, *Anopheles stphensi*," *Parasitology*, vol. 108, no. 4, pp. 383–388, 1994.
- [107] J. S. Kayembe, K. M. Taba, K. Ntumba et al., "In vitro antimalarial activity of 20 quinones isolated from four plants used by traditional healers in the Democratic Republic of Congo," Journal of Medicinal Plants Research, vol. 4, no. 11, pp. 991–994, 2010.
- [108] J. S. Kayembe, K. M. Taba, K. Ntumba et al., "In vitro antimalarial activity of eleven terpenes isolated from Ocimum gratissimum and Cassia alata leaves screening of their binding affinity with Haemin," Journal of Plant Studies, vol. 1, no. 2, pp. 168–172, 2012.
- [109] B. Y. Vigbedor, A. S. Osafo, A. G. Ben et al., "In vitro antimalarial activity of the ethanol extracts of Afzelia africana and Cassia alata commonly used as herbal remedies for malaria in Ghana," International Journal of Novel Research in Life Science, vol. 2, no. 6, pp. 10–16, 2015.
- [110] J. Anbu, M. Anita, R. Sathiya et al., "In Vitro Anthelmintic activity of leaf ethanolic extract of Cassia alata and Typha angustifolia," MSRUAS-SASTech Journal, vol. 14, no. 2, pp. 41–44, 2013.
- [111] A. Angelina, M. Hanafi, F. Suyatna et al., "Antiviral effect of sub fraction *Cassia alata* leaves extract to dengue virus Serotype-2 strain new guinea C in human cell line Huh-7 it-1," *IOP Conference Series: Earth and Environmental Science*, vol. 101, Article ID 012004, 2017.
- [112] J. A. Owoyale, G. A. Olatunji, and S. O. Oguntoye, "Antifungal and antibacterial activities of an alcoholic extract of *S. alata* leaves," *Journal of Applied Science and Environment*, vol. 9, no. 3, pp. 105–107, 2005.
- [113] A. H. Gilani and A. U. Rahman, "Trends in ethnopharmacology," *Journal of Ethnopharmacology*, vol. 100, pp. 43–49, 2005.
- [114] U. A. Essiett and I. E. Bassey, "Comparative phytochemical screening and nutritional potentials of the flowers (petals) of S. alata (l) roxb, Senna hirsuta (l.) Irwin and barneby, and Senna obtusifolia (l.)," Journal of Applied Pharmaceutical Science, vol. 3, no. 8, pp. 097–101, 2013.
- [115] I. Abdulwaliyu, S. O. Arekemase, S. Bala et al., "Nutritional properties of *S. alata* Linn leaf and flower," *International*

- Journal of Modern Biological Medicine, vol. 4, no. 1, pp. 1–11, 2013
- [116] A. P. Grollman and D. M. Marcus, "Global hazards of herbal remedies: lessons from Aristolochia," *EMBO Representation*, vol. 17, pp. 619–625, 2016.
- [117] A. E. Ugbogu, E. Okezie, C. Uche-Ikonne et al., "Toxicity evaluation of the aqueous stem extracts of *S. alata* in wistar rats," *American Journal of Biomedical Research*, vol. 4, no. 4, pp. 80–86, 2016.
- [118] I. Igbe and E. Osaze, "Toxicity profile of aqueous extract of *Cassia alata* flower in wistar rats," *Journal of Pharmacy and Bioresources*, vol. 13, no. 2, pp. 92–102, 2016.
- [119] S. Roy, B. Ukil, and L. M. Lyndem, "Acute and sub-acute toxicity studies on the effect of *S. alata* in swiss albino mice," *Cogent Biology*, vol. 2, no. 1, p. 127, 2016.
- [120] A. Rezaei, A. Farzadfard, A. Amirahmadi, M. Alemi, and M. Khademi, "Diabetes mellitus and its management with medicinal plants: a perspective based on Iranian research," Journal of Ethnopharmacology, vol. 175, pp. 567–616, 2015.
- [121] F. C. Onwuliri, "Antimicrobial studies of the extracts of *Acalypha wllkesiana* L. on microorganisms associated with wound and skin infections," *West African Journal of Biological Science*, vol. 15, pp. 15–19, 2004.