



ELSEVIER

Contents lists available at ScienceDirect

Data in Brief

journal homepage: www.elsevier.com/locate/dib



Data Article

Data on energy and economic evaluation and microbial assessment of anaerobic co-digestion of fruit rind of *Telfairia occidentalis* (Fluted pumpkin) and poultry manure



CrossMark

S.O. Dahunsi ^{a,b,*}, A. Olayanju ^{b,c}, J.O. Izebere ^b, A.P. Oluyori ^d

^a Faculty of Environment and Labour Safety, Ton Duc Thang University, Ho Chi Minh City, Vietnam

^b Biomass and Bioenergy Group, Environment and Technology Research Cluster, Landmark University, Omu-Aran, Kwara State, Nigeria

^c Department of Agricultural and Biosystems Engineering, Landmark University, Omu-Aran, Kwara State, Nigeria

^d Physical Sciences Department, Landmark University, Omu-Aran, Kwara State, Nigeria

ARTICLE INFO

Article history:

Received 9 June 2018

Received in revised form

20 September 2018

Accepted 24 September 2018

Keywords:

Biogas

Biomass

Economics

Energy

Fluted pumpkin

Microorganisms

ABSTRACT

The data described in this article was obtained in an experiment designed for the generation of biogas from the anaerobic co-digestion of *Telfairia occidentalis* (Fluted pumpkin) fruit rind and poultry manure both of which currently constitute an environmental nuisance in the localities where they are found. The data presented in this article is on the use of combined heat and power (CHP) system to assess the energy and economic feasibility of applying thermo-alkali pretreatment procedures to one of the substrates (Fluted pumpkin) prior to anaerobic digestion. Also, the microbial characterization and succession pattern of important microbes during the anaerobic digestion process was evaluated and the data reported in this paper.

© 2018 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

* Corresponding author at: Faculty of Environment and Labour Safety, Ton Duc Thang University, Ho Chi Minh City, Vietnam.
E-mail address: dahunsi.olatunde.samuel@tdt.vn (S.O. Dahunsi).

Specifications table

| | |
|----------------------------|--|
| Subject area | Microbiology and Biotechnology |
| More specific subject area | Environmental Biotechnology |
| Type of data | Tables |
| How data was acquired | Combined Heat and Power (CHP) System, Analytical Profile Index (API) kits (BioMerieux, Leon, France) |
| Data format | Analysed |
| Experimental factors | Produced thermal energy, produced electrical energy, thermal energy gain, thermal energy requirement, net thermal energy, electrical energy gain, electrical energy requirement, net electrical energy |
| Experimental features | Energy and Economic evaluation of anaerobic co-digestion of pre-treated and non-pretreated fruit rind of <i>Telfairia occidentalis</i> (Fluted Pumpkin) and Poultry Manure |
| Data source location | Omoo-Aran, Kwara State |
| Data accessibility | The data is available within the article body |

Values of the data

- The data presented in this article reveals the energy and economic evaluation of the anaerobic co-digestion of fruit rind of *Telfairia occidentalis* (Fluted Pumpkin) and Poultry manure for biogas generation
 - The data will serve as a precursor for further research on the economic assessment of biomass pretreatment prior to anaerobic digestion processes
 - The data give further exposure on the necessity and feasibility of pretreatment of biomass prior to anaerobic digestion.
 - More robust heat and power systems can be used to further explore the generated data from this study in order to apply the processes in industrial scale experiments.
-

1. Data

The combined heat and power (CHP) system was used to assess the energy balance and the economic feasibility of applying thermal and alkaline pre-treatment to *T. occidentalis* fruit rind using a 50 and 30% thermal and electrical efficiencies respectively ([Table 1](#)). Therefore, to determine the thermal energy requirement (TER) for thermal and alkaline pre-treatments of *T. occidentalis* fruit rind, the energy needed to raise the temperature of 35 g TS L⁻¹ *T. occidentalis* fruit rind mixture from 25 to 55 °C was determined using the specific heat of water i.e. 4.18 kJ kg⁻¹ °C⁻¹ in order to evaluate the specific heat of the mixture while neglecting heat loss [[1–3](#)].

To assess the electrical energy, only the electric energy used for the substrate mixing was considered neglecting the energy used during mechanical treatment since this was also done for the experiment without thermal and alkaline pre-treatment [[4](#)]. [Table 2](#) shows the heat balance of different biomass previously anaerobically digested with thermal and alkaline pre-treatments procedures [[5–9](#)].

In the co-digestion of *Telfairia occidentalis* fruit rind and poultry manure, various aerobic and anaerobes bacteria, fungi and methanogens were isolated and characterized ([Table 3](#)).

Table 1Energy and economic evaluation for the anaerobic co-digestion of *Telfairia occidentalis* fruit rind and poultry manure.

| Energy parameters | Experiment A | Experiment B | Experiment C |
|---|---------------------|---------------------|---------------------|
| Produced electrical and thermal energy from combined heat and power (CHP) | 1785 ± 0.01 | 1699 ± 0.02 | 1155 ± 0.02 |
| Produced thermal energy (kWh t ⁻¹ TS) | 1645 ± 0.02 | 1547 ± 0.01 | 498 ± 0.01 |
| Produced electrical energy (kWh t ⁻¹ TS) | 770 ± 0.01 | 563 ± 0.02 | 340 ± 0.02 |
| Thermal balance | | | |
| *Thermal energy gain (kWh t ⁻¹ TS) | 1147 ± 0.01 | 1049 ± 0.03 | – |
| Thermal energy requirement (kWh t ⁻¹ TS) | 1088 ± 0.02 | 1109 ± 0.03 | – |
| Thermal energy requirement with 80% of heat recovery (kWh t ⁻¹ TS) | 218 ± 0.02 | 210 ± 0.01 | – |
| #Net thermal energy (kWh t ⁻¹ TS) | 59 ± 0.02 | –60 ± 0.02 | – |
| Net thermal energy with 80% of heat recovery (kWh t ⁻¹ TS) | –929 ± 0.02 | –839 ± 0.03 | – |
| Electrical balance | | | |
| [§] Electrical energy gain | 430 ± 0.01 | 223 ± 0.02 | – |
| Energy for mixing during pretreatment | – | – | – |
| Net electrical energy | 430 ± 0.01 | 223 ± 0.01 | – |
| Economic evaluation | | | |
| Cost of NaOH (€ t ⁻¹ TS) | – | – | – |

Remark:*=difference of thermal energies produced by the pretreated experiment minus the untreated; #=difference between the thermal energy gain and the thermal energy requirement for the thermo-alkaline pretreatment; a [§]=difference of electricity energies produced by pretreated experiment minus the untreated.

2. Experimental design, materials and methods

2.1. Materials and method

Data was obtained from the evaluation of pretreatment application to fruit rind of *Telfairia occidentalis* and the possibility of gaining back the investment (obtaining of chemicals and heat) into the pretreatment procedure through the sale of additional energy gained.

2.2. Experimental design

A simple computational equation was used to first determine the thermal energy required (TER) in kWh t⁻¹ TS for raising the temperature of one ton TS of *T. occidentalis* fruit rind from 25 to 55 °C during pre-treatment [14–16].

2.3. Microbial enumeration

The aerobic organisms (Bacteria and fungi) associated with the fermenting substrates were isolated and enumerated weekly using standard methods [17–19]. Facultative anaerobes were serially isolated using specialized media in an anoxic condition at 37 °C for 5 to 7 days as earlier reported [20,21]. Confirmation of the presumptive isolates was done with corresponding rapid Analytical Profile Index (API) kits [22] while a basal medium was used for identifying methanogens [23,24].

Table 2

Energy balances of thermal and thermo-chemical pretreatment procedures as applied to different substrates.

| Substrate | Condition of pretreatment | Increase in methane yield ($\text{m}^3 \text{ t}^{-1} \text{ TS}$)/operation mode | Biogas conversion | Surplus thermal energy ($\text{kWh t}^{-1} \text{ TS}$) | Thermal pretreatment requirements ($\text{kWh t}^{-1} \text{ TS}$) | Net heat energy ($\text{kWh t}^{-1} \text{ TS}$) | References |
|--|--|---|--------------------------------|---|--|--|---------------|
| <i>Telfairia occidentalis</i> fruit rind | Thermo-alkaline (55 °C; 4% NaOH (w/w); 24 h) Solid load: 35 g TS L ⁻¹ | 40/Batch mode | CHP: 35% electricity; 50% heat | 1147 | 1088 | 59 | Current study |
| | Thermo-alkaline (55 °C; 4% KOH (w/w); 24 h) Solid load: 35 g TS L ⁻¹ | 35/Batch mode | CHP: 35% electricity; 50% heat | 1049 | 1109 | -60 | Current study |
| <i>Tithonia diversifolia</i> shoot | Thermo-alkaline (55 °C; 4% NaOH (w/w); 24 h) Solid load: 35 g TS L ⁻¹ | 53/Batch mode | CHP: 35% electricity; 50% heat | 1176 | 1068 | 108 | [10] |
| | Thermo-alkaline (55 °C; 4% KOH (w/w); 24 h) Solid load: 35 g TS L ⁻¹ | 30/Batch mode | CHP: 35% electricity; 50% heat | 862 | 1150 | -288 | [10] |
| Peanut hull | Thermo-alkaline (55 °C; 4% NaOH (w/w); 24 h) Solid load: 35 g TS L ⁻¹ | 70/Batch mode | CHP: 35% electricity; 50% heat | 761 | 1173 | -412 | [11] |
| Sunflower stalks | Thermo-alkaline (55 °C; 4% NaOH (w/w); 24 h) Solid load: 35 g TS L ⁻¹ | 36/Continuous mode | CHP: 35% electricity; 50% heat | 185 | 1034 | -849 | [12] |
| | Thermo-alkaline (55 °C; 4% NaOH (w/w); 24 h) Solid load: 50 g TS L ⁻¹ | 36/Continuous mode | CHP: 35% electricity; 50% heat | 185 | 733 | -548 | [12] |
| | thermo-alkaline (55 °C; 4% NaOH (w/w TS); 24 h) Solid load: 200 g TS L ⁻¹ | 36/Continuous mode | CHP: 35% electricity; 50% heat | 185 | 210 | -25 | [12] |
| | Thermo-alkaline (55 °C; 4% NaOH (w/w); 24 h) Solid load: 50 g TS L ⁻¹ 80% of heat recovery from pretreatment | 36/Continuous mode | CHP: 35% electricity; 50% heat | 185 | 147 | 38 | [12] |
| | Thermal (170 °C; 1 h) Solid load: 50 g TS L ⁻¹ | 32/Batch mode | CHP: 35% electricity; 50% heat | 161 | 3535 | -3375 | [6] |
| | Thermal (170 °C; 1 h) Solid load: 200 g TS L ⁻¹ | 32/Batch mode | CHP: 35% electricity; 50% heat | 161 | 1010 | -849 | [6] |
| Sunflower Oil Cake | Thermal (170 °C; 1 h) Solid load: 200 g TS L ⁻¹ Thermal (170 °C; 1 h) Solid load: 200 g TS L ⁻¹ 80% of heat recovery from pretreatment | 32/Batch mode | CHP: 35% electricity; 50% heat | 161 | 152 | 9 | [6] |

| | | | | | | |
|------------------------|--|--|------------|------------|--------------|------|
| Ensiled Sorghum Forage | Thermo-alkaline (100 °C; 30 min, 10% NaOH w/w) Solid load: 160 g TS L ⁻¹ Thermo-alkaline (100 °C; 30 min, 10% NaOH w/w) Solid load: 160 g TS L ⁻¹ 80% of heat recovery from Pretreatment | 92/Batch mode CHP: 40% electricity; 41% heat CHP: 40% electricity; 41% heat | 378 378 | 547 109 | – 169 269 | [13] |
| Wheat straw | Thermo-alkaline (100 °C; 30 min, 10% NaOH w/w) Solid load: 160 g TS L ⁻¹ Thermo-alkaline (100 °C; 30 min, 10% NaOH w/w) Solid load: 160 g TS L ⁻¹ 80% of heat recovery from Pretreatment | 137/Batch mode CHP: 40% electricity; 41% heat CHP: 40% electricity; 41% heat | 577 577 | 547 109 | 30 468 | [13] |
| Microalgae | Thermal (75 °C; 15 min) Solid load: 11.7 g TS L ⁻¹ 85% of heat recovery from Pretreatment | 32/Batch mode 100% heat conversion | 316 | 458 | – 142 | [7] |
| | Thermal (75 °C; 15 min) Solid load: 20 g TS L ⁻¹ 85% of heat recovery from Pretreatment | 32/Batch mode 100% heat conversion | 316 | 268 | 48 | [7] |
| | Thermal (75 °C; 15 min) Solid load: 30 g TS L ⁻¹ 85% of heat recovery from Pretreatment | 32/Batch mode 100% heat conversion | 316 | 173 | 143 | [7] |

Table 3

Microbial evaluation and succession in the anaerobic co-digestion of *Telfairia occidentalis* fruit rind+poultry manure.

| Day | Aerobes (Cfu/ml) | | Fungi (Cfu/ml) | | Anaerobes (Cfu/ml) | | Methanogens (Cfu/ml) | |
|-----|---|----------------------|--|-------------------|--|----------------------|---|----------------------|
| | Organism | TAPC | Organism | TFC | Organism | TPC | Organism | TPC |
| 0 | <i>Bacillus</i> sp. <i>Serratia</i> sp. <i>Pseudomonas aeruginosa</i> <i>Proteus</i> sp. | 2.3×10^{10} | <i>Aspergillus niger</i> <i>Aspergillus flavus</i> <i>Rhizopus</i> sp. <i>Mucor</i> sp. <i>Penicillium</i> sp. | 1.0×10^8 | <i>Fusobacterium</i> sp. <i>Bacteroides</i> sp. <i>Clostridium</i> sp. <i>Porphyromonas</i> sp. | 1.2×10^{10} | <i>Methanoscincales</i> sp. <i>Methanobacteriales</i> sp. <i>Methanomicrobiales</i> sp. <i>Aminobacteria</i> sp. | 1.2×10^{10} |
| 6 | <i>Bacillus</i> sp. <i>Serratia</i> sp. <i>Pseudomonas aeruginosa</i> <i>Proteus</i> sp. | 1.4×10^8 | <i>Aspergillus niger</i> <i>Aspergillus flavus</i> <i>Rhizopus</i> sp. <i>Mucor</i> sp. <i>Penicillium</i> sp. | 1.2×10^8 | <i>Fusobacterium</i> sp. <i>Bacteroides</i> sp. <i>Clostridium</i> sp. <i>Porphyromonas</i> sp. | 1.0×10^6 | <i>Methanoscincales</i> sp. <i>Methanobacteriales</i> sp. <i>Methanomicrobiales</i> sp. <i>Aminobacteria</i> sp. | 1.0×10^8 |
| 12 | Nil | Nil | <i>Aspergillus niger</i> <i>Aspergillus flavus</i> <i>Rhizopus</i> sp. <i>Mucor</i> sp. <i>Penicillium</i> sp. | 1.0×10^3 | <i>Fusobacterium</i> sp. <i>Bacteroides</i> sp. <i>Clostridium</i> sp. <i>Porphyromonas</i> sp. | 1.0×10^4 | <i>Methanoscincales</i> sp. <i>Methanobacteriales</i> sp. <i>Methanomicrobiales</i> sp. <i>Aminobacteria</i> sp. | 1.0×10^5 |
| 18 | <i>Bacillus</i> sp. | 1.0×10^2 | <i>Aspergillus niger</i> | 1.0×10^2 | <i>Fusobacterium</i> sp. <i>Clostridium</i> sp. <i>Porphyromonas</i> sp. | 1.3×10^{10} | <i>Methanoscincales</i> sp. <i>Methanobacteriales</i> sp. <i>Methanomicrobiales</i> sp. <i>Aminobacteria</i> sp. | 1.0×10^{10} |
| 24 | <i>Bacillus</i> sp. | 1.0×10^2 | <i>Aspergillus niger</i> | 1.0×10^2 | <i>Fusobacterium</i> sp. <i>Clostridium</i> sp. <i>Porphyromonas</i> sp. | 1.2×10^3 | <i>Methanoscincales</i> sp. <i>Methanobacteriales</i> sp. <i>Methanomicrobiales</i> sp. <i>Aminobacteria</i> sp. | 1.7×10^{10} |
| 30 | <i>Bacillus</i> sp. | 1.0×10^2 | <i>Aspergillus niger</i> | 1.0×10^2 | <i>Fusobacterium</i> sp. <i>Clostridium</i> sp. | 1.2×10^2 | <i>Methanoscincales</i> sp. <i>Methanobacteriales</i> sp. <i>Methanomicrobiales</i> sp. <i>Aminobacteria</i> sp. | 2.7×10^{12} |

Remark: TAPC=Total aerobic plate count; TFC=Total fungal count; TPC=Mean Plate Count.

2.4. Statistical data analysis

The paired sample t-tests were conducted to determine the significant difference in the means of three replicates.

Acknowledgment

The author is grateful to Ton Duc Thang University, Ho Chi Mihn City, Vietnam for funding this research.

Transparency document. Supporting information

Transparency data associated with this article can be found in the online version at <https://doi.org/10.1016/j.dib.2018.09.065>.

References

- [1] G.D. Zupancic, M. Ros, Heat and energy requirements in thermophilic anaerobic sludge digestion, *Renew. Energy* 28 (2003) 2255–2267.
- [2] B.R. Dhar, G. Nakhla, M.B. Ray, Techno-economic evaluation of ultrasound and thermal pretreatments for enhanced anaerobic digestion of municipal waste activated sludge, *Waste Manag.* 32 (2012) 542–549.
- [3] J. Zabranska, M. Dohanyos, P. Jenicek, J. Kutil, Disintegration of excess activated sludge – evaluation and experience of full-scale applications, *Water Sci. Technol.* 53 (2006) 229–236.
- [4] S. Menardo, G. Airoldi, P. Balsari, The effect of particle size and thermal pretreatment on the methane yield of four agricultural by-products, *Bioresour. Technol.* 104 (2012) 708–714.
- [5] F. Fdz-Polanco, V. Velazquez, I. Perez-Elvira, C. Casas, D. del Barrio, F.J. Cantero, F. Fdz-Polanco, P. Rodriguez, L. Panizo, J. Serra, P. Rouge, Continuous thermal hydrolysis and energy integration in sludge anaerobic digestion plants, *Water Sci. Technol.* 57 (8) (2008) 1221–1226.
- [6] F. Monlau, E. Latrille, A.C. Da Costa, J. Steyer, H. Carrère, Enhancement of methane production from sunflower oil cakes by dilute acid pretreatment, *Appl. Energy* 102 (2013) 1105–1113.
- [7] F. Passos, J. Garcia, I. Ferrer, Impact of low temperature pretreatment on the anaerobic digestion of microalgal biomass, *Bioresour. Technol.* 138 (2013) 79–86.
- [8] D.J. Schell, J. Farmer, M. Newman, J.D. McMillan, Dilute-sulfuric acid pretreatment of corn stover in the pilot-scale reactor: investigation of yields, kinetics, and enzymatic digestibilities of solids, *Appl. Biochem. Biotechnol.* 105–108 (2003) 69–85.
- [9] A.A. Modenbach, S.E. Nokes, The use of high-solids loading in biomass pretreatment – a review, *Biotechnol. Bioeng.* 109 (2012) 1430–1442.
- [10] S.O. Dahunsi, O. Oranusi, V.E. Efeovbokhan, Bioconversion of *tithoniadiversifolia* (Mexican Sunflower) and poultry droppings for energy generation: optimization, mass, and energy balances, and economic benefits, *Energy Fuels* 31 (2017) 5145–5157.
- [11] S.O. Dahunsi, O. Oranusi, V.E. Efeovbokhan, Cleaner energy for cleaner production: modeling and optimization of biogas generation from *Carica papaya* (Pawpaw) fruit peels, *J. Clean. Prod.* 156 (2017) 19–29.
- [12] F. Monlau, C. Sambusiti, N. Antoniou, A. Barakat, A. Zabaniotou, A new concept for enhancing energy recovery from agricultural residues coupling anaerobic digestion and pyrolysis process, *Appl. Energy* 148 (2015) 32–38.
- [13] C. Sambusiti, E. Ficara, F. Malpei, J.P. Steyer, H. Carrere, Benefit of sodium hydroxide pretreatment of ensiled sorghum forage on the anaerobic reactor stability and methane production, *Bioresour. Technol.* 144 (2013) 149–155.
- [14] S.O. Dahunsi, O. Oranusi, V.E. Efeovbokhan, Optimization of pretreatment, process performance, mass, and energy balance in the anaerobic digestion of *Arachis hypogaea* (Peanut) hull, *Energy Conv. Manag.* 139 (2017) 260–275.
- [15] S.O. Dahunsi, O. Oranusi, V.E. Efeovbokhan, Anaerobic mono-digestion of *Tithonia diversifolia* (Wild Mexican sunflower), *Energy Conv. Manag.* 148 (2017) 128–145.
- [16] S.O. Dahunsi, O. Oranusi, V.E. Efeovbokhan, Pretreatment optimization, process control, mass, and energy balances and economics of anaerobic co-digestion of *Arachis hypogaea* (Peanut) hull and poultry manure, *Bioresour. Technol.* 241 (2017) 454–464.
- [17] A. Tsuneyo, *Pictorial Atlas of Soil for Seed Fungi: Morphologies of Cultural Fungi for the Key to Species*, Third Edition, CRC Press, USA, 2010.
- [18] S.O. Dahunsi, O. Oranusi, J.B. Owolabi, V.E. Efeovbokhan, Comparative biogas generation from fruit peels of Fluted Pumpkin (*Telfairia occidentalis*) and its optimization, *Bioresour. Technol.* 221 (2016) 517–525.
- [19] S.O. Dahunsi, O. Oranusi, J.B. Owolabi, V.E. Efeovbokhan, Mesophilic anaerobic co-digestion of poultry droppings and *Carica papaya* peels: modelling and process parameter optimization study, *Bioresour. Technol.* 216 (2016) 587–600.
- [20] S.O. Dahunsi, O. Oranusi, J.B. Owolabi, V.E. Efeovbokhan, Synergy of Siam weed (*Chromolaena odorata*) and poultry manure for energy generation: effects of pretreatment methods, modeling and process optimization, *Bioresour. Technol.* 225 (2016) 409–417.

- [21] T.A. Ayandiran, A.A. Ayandele, S.O. Dahunsi, O.O. Ajala, Microbial assessment and prevalence of antibiotic resistance in polluted Oluwa River, Nigeria, *Egypt J. Aqua Res.* 40 (2014) 291–299.
- [22] T.A. Ayandiran, S.O. Dahunsi, Microbial evaluation and occurrence of antidiug multi-resistant organisms among the indigenous Clarias species in River Oluwa, Nigeria, *J. King Saud. Univ- Sci.* 29 (2017) 96–105.
- [23] S. Ghosh, P. Jha, A.S. Vidyarthi, Unravelling the microbial interactions in coal organic fermentation for generation of methane—a classical to metagenomic approach, *Int. J. Coal Geol.* 125 (2014) 36–44.
- [24] M. Stieglmeier, R. Wirth, G. Kminek, C. Moissl-Eichinger, Cultivation of anaerobic and facultatively anaerobic bacteria from spacecraft-associated clean rooms, *Appl. Environ. Microbiol.* 75 (2009) 3484–3491.